Overview of Lecture Series

1. Determining the molecular identity of receptors

Getting the first clone

Classical approach Expression cloning

Isoforms & Evolutionary Precursors PCR

2. Determining the stoichiometry of receptors

Selective Tagging

Constraining Stoichiometry

3. Determining the structure of receptors

Membrane Topology

Hydropathy Plots Engineered Reporter Sites

<u>3-Dimensional Structure</u>

Electron Cryomicrocopy X-ray crystallography

Methodological Approaches To Study Receptors

Lecture #1

Determining The Molecular Identity of Receptors



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Overview

Phase 1: <u>Getting The First Clone</u>

Classical Approach e.g. nAChRs
Positional Cloning e.g. K-channels
Expression Cloning e.g. iGluRs

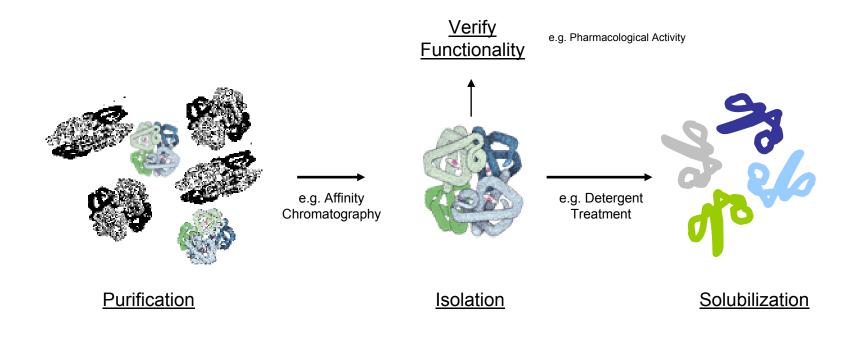
Phase 2: Isoforms & Evolutionary Precursors

Low-Stringency Hybridization
Polymerase Chain Reaction
Searching Databases

Getting the First Clone: Classical Approach

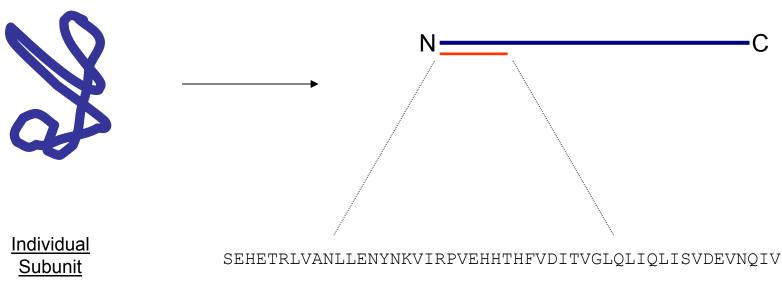
Step 1: Purify From A Source Rich In Protein of Interest

e.g. Na channel or nAChR: Electric organ of electric eel



Getting the First Clone: Classical Approach

Step 2: Obtain Partial Sequence Of Protein



(e.g. Chemical sequencing of N-terminal AAs

Approx. first 50 residues)

e.g. α-subunit of nAChR, Torpedo californica

Getting the First Clone: Classical Approach

Step 3: Synthesis Of Oligonucleotide Probes

Partial Protein Sequence

SEHETRLVANLLENYNKVIRPVEHHTHFVDITVGLQLIQLISVDEVNQIV

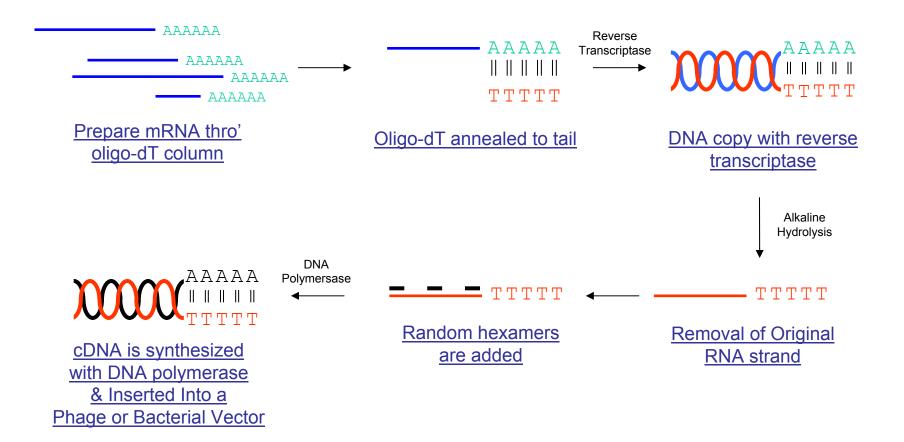
e.g. α -subunit of nAChR, Torpedo californica

Nucleotide Sequence To Generate Probes For cDNA Library

ATCTCTTCACTAGAAAAGAGCTGAACACAGAAGTCCAGAAGATCTAACAAGTTCATCGTTTAGTTATTAGAAGTG GCAGATTTGCTTGAAAAGCCCAATTATTGAAAGCTGAAGAATGATTCTGTGCAGTTATTGGCATGTAGGGTTGGTG

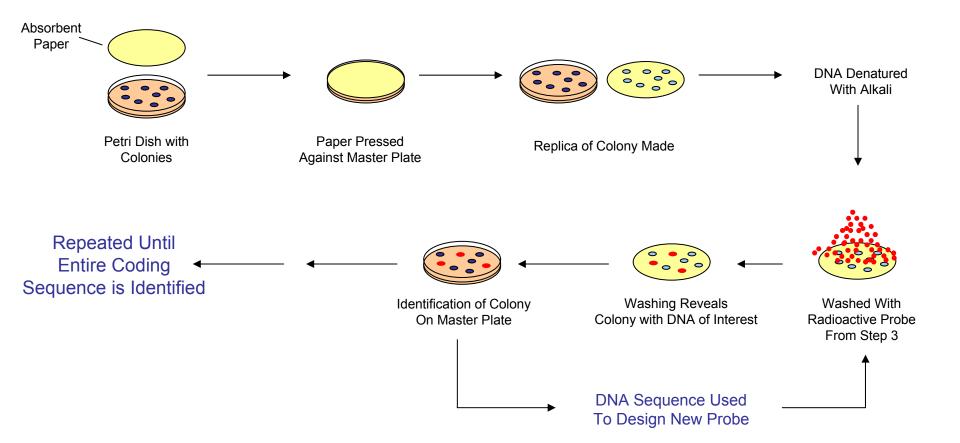
Getting the First Clone: Classical Approach

Step 4: Generating a cDNA Library



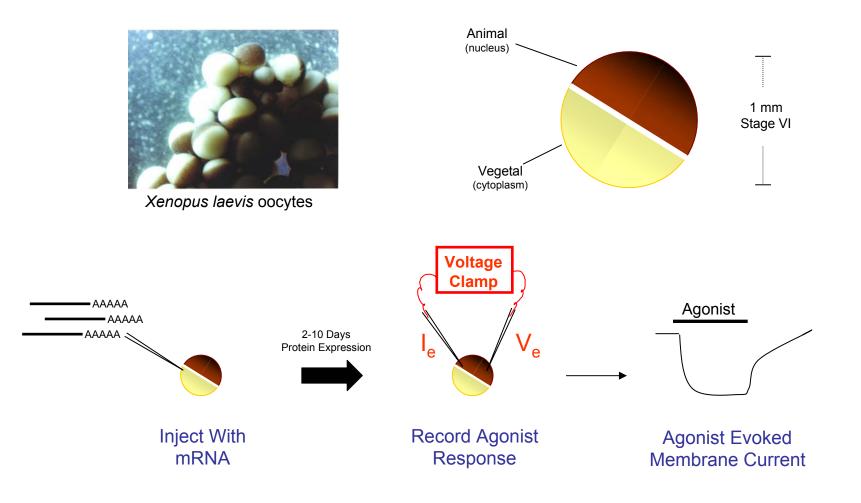
Getting the First Clone: Classical Approach

Step 5: Screening cDNA Library



<u>Getting the First Clone</u>: Classical Approach

Step 6: Verify Function In Heterologous Expression System



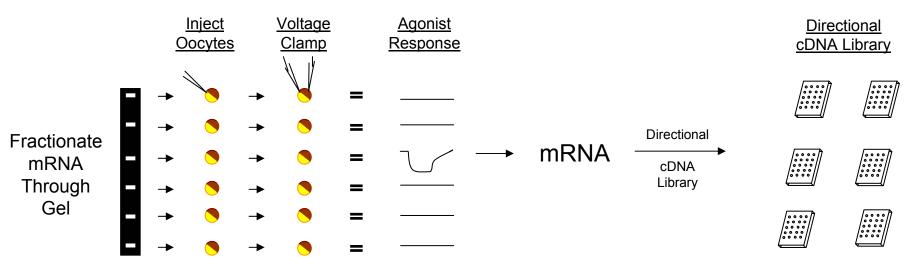
Getting the First Clone: Classical Approach

Benefits & Disadvantages

- Slow, requires protein chemistry, multi-disciplinary
- Limited, Needs Source Rich In Protein
- Successfully employed however e.g. nAChRs, GABA_A & Glycine, Na channels

<u>Getting the First Clone</u>: Expression Cloning

Step 1: Construct cDNA Library Enriched For Receptor Activity

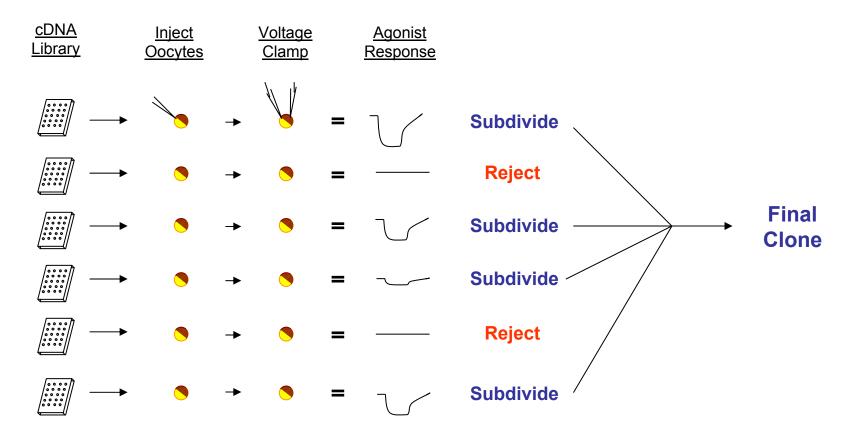


Consisting of Several Independent Sub-Libraries

e.g. Source of Rat Forebrain, First iGluR clone Hollmann *et al* (1989) Nature 342, 643-648

Getting the First Clone: Expression Cloning

Step 2: Screening of Sub-Libraries



<u>Getting the First Clone</u>: Expression Cloning

Benefits & Disadvantages

- Faster, neither protein chemistry nor sequence info required
- Works if single mRNA makes a functional product



Clone may represent an enzyme or protein that aids expression of receptor

What Have We Learned So Far ?

1. Getting the first clone

Classical approach Expression cloning

What's Next?

2. Isoforms & Evolutionary Precursors

Polymerase Chain Reaction

Isoforms & Evolutionary Precursors: PCR

Objective: Amplify Many Copies Of Homologous Sequences

Step 1: Setting Up Reaction Conditions

Requires Sequence Information Of 2 Regions of DNA

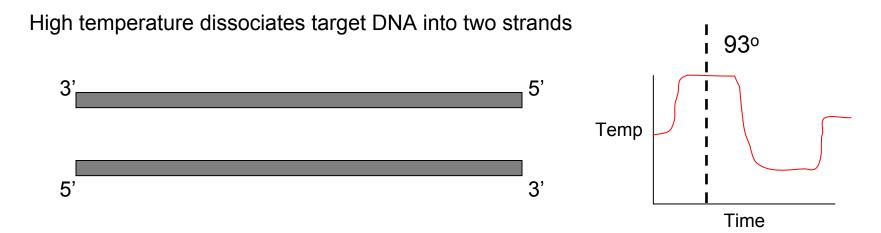
Two Primers Are Synthesized (15-20 nucleotides)

Primers Are 200-2000 bps apart allowing 3' ends to point to each other

Isoforms & Evolutionary Precursors: PCR

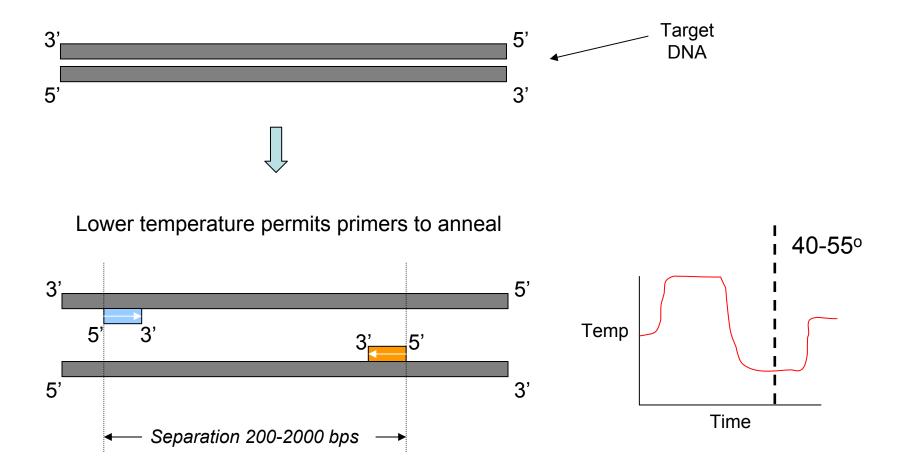
Step 2: Denaturation of Target DNA





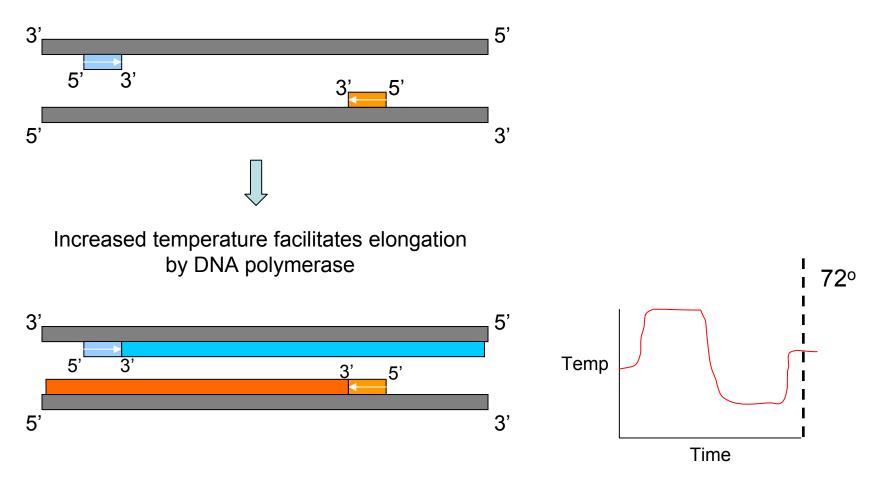
Isoforms & Evolutionary Precursors: PCR

Step 3: Annealing Primers To Target DNA



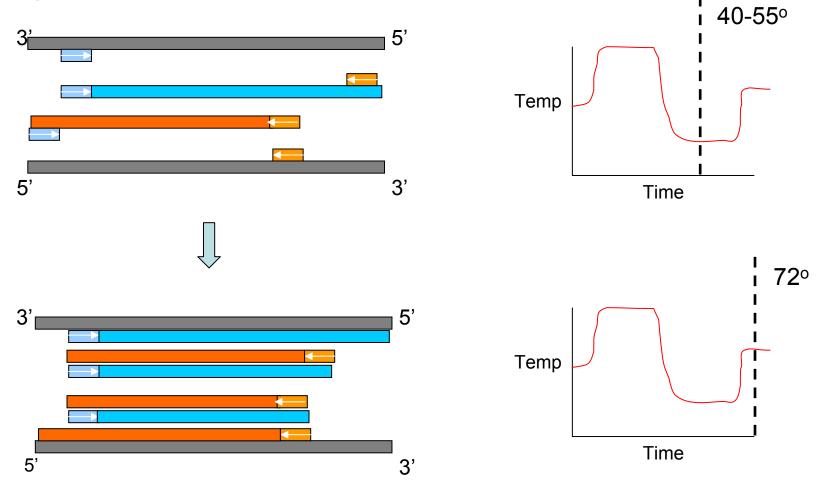
Isoforms & Evolutionary Precursors: PCR

Step 4: Elongation of DNA by DNA Polymerase



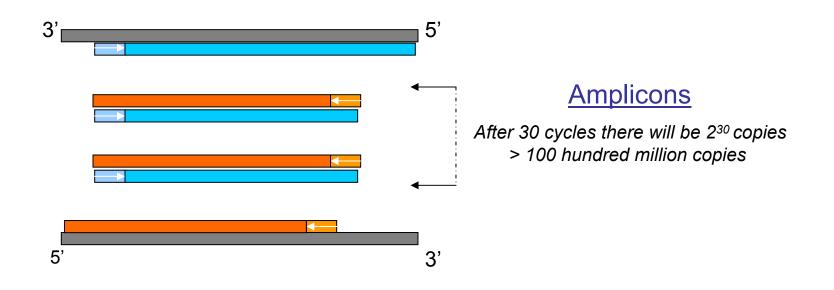
Isoforms & Evolutionary Precursors: PCR

Step 5: Second PCR Cycle Further Amplifies Copy Number



Isoforms & Evolutionary Precursors: PCR

Step 6 onwards: Process is Repeated Many Times



What Have We Learned?

1. Getting the first clone

Classical approach Expression cloning

2. Isoforms & Evolutionary Precursors

Polymerase Chain Reaction

Further Reading

1. Snutch, T.P. (1988) The use of Xenopus oocytes to probe synaptic communication. TiNS, 11, 250-256.

2. Hille, B. (1996) Ion channels of excitable membranes; Structure of channel proteins Chapter 13, 3rd Edition, Sinauer Associates, Inc.

3. Simonsen, H. & Lodish, H.F. (1994) Cloning by function: expression cloning in mammalian cells. TiPS, 15, 437-441.

4. Hollmann *et al* (1989) Cloning by functional expression of a member of the glutamate receptor family. Nature, 342, 643-648.